

Impact of Glyphosate on the Development of Type-2 Diabetes in Adipose Tissue: Role of Insulin-Like Growth Factor 1 and Tumor Necrosis Factor Alpha Expression

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Abstract

Glyphosate, a commonly used herbicide in agricultural and residential settings, has sparked concerns regarding its potential health impacts. Despite numerous studies exploring potential associations between glyphosate exposure and diabetes, the precise mechanisms remain unclear. Tumor Necrosis Factor-alpha (TNF- α) and Insulin-Like Growth Factor 1 (IGF-1) are implicated in insulin resistance and pancreatic beta cell dysfunction, playing crucial roles in diabetes pathogenesis, especially Type 2 diabetes. This study aimed to assess the effect of glyphosate on TNF- α and IGF-1 expression in male Wistar rats. The results revealed a dose-dependent increase ($p < 0.05$) in TNF- α and IGF-1 expression in adipose tissue following glyphosate exposure compared to the control group. These findings suggest that glyphosate exposure may contribute to the development of diabetes by altering the expression of IGF-1 and TNF- α .

Keywords: Herbicide, IGF-1, Innovative Technology, Novel Method, Type-2 Diabetes, TNF- α .

Introduction

Glyphosate, an organophosphate compound widely used as an herbicide, exerts its action by inhibiting crucial plant enzymes involved in metabolic processes. Acute exposure studies on male Wistar rats have revealed intriguing molecular effects, including increased aromatase mRNA and protein expression, alongside a decrease in normal sperm count attributed to reduced levels of histone-1 and protamine-1 [1]. Roundup, a glyphosate-based herbicide, has also been investigated for its impact on testicular cells, showing potential necrotic damage to Leydig cells following a 48-hour exposure period [2].

Moreover, high exposure to glyphosate-based herbicides like Roundup has been linked

to Leydig cell necrosis in vivo [3]. Shifting focus to diabetes, characterized by impaired regulation of blood glucose levels leading to hyperglycemia, insulin deficiency plays a pivotal role, manifesting in symptoms such as polyuria, polydipsia, weight loss, and blurred vision [4]. Autoimmune destruction of pancreatic beta cells, responsible for insulin production, underlies this deficiency, resulting in hyperglycemia and subsequent diabetes-related complications [5].

Insulin-like Growth Factor 1 (IGF-1), primarily involved in regulating bone formation, is predominantly synthesized by osteoblasts, the body's bone-forming cells. Notably, IGF-1 also influences systemic hormone effects on bone formation and has been implicated in increasing osteoclast

formation from osteoclast precursors in mice. It is a crucial growth factor stored in the bone matrix, aiding in bone resorption and subsequent coupling of bone formation processes [6].

Tumor Necrosis Factor-alpha (TNF- α), produced by macrophages, plays a pivotal role in tumor cell destruction [7]. However, the precise impact of glyphosate on inflammatory markers remains elusive, despite previous research indicating its association with increased anxiety levels and cognitive deficits in diabetic rats [8-27]. Leveraging our team's expertise and extensive research experience, we aim to fill this gap by analyzing the effect of glyphosate exposure on the expression of IGF-1 and TNF- α in the liver.

Materials and Methods

Chemicals and Reagents

All chemicals and reagents utilized in this study were procured from reputable sources.

Animal Ethics and Maintenance

The experimental procedures involving animals were ethically approved and maintained as per the standard procedure.

Experimental Design

Adult male Wistar albino rats were divided into four groups, each comprising six animals. Group I served as the normal control and was fed a standard diet with access to drinking water. Groups II, III, and IV were orally administered glyphosate dissolved in water at doses of 50 mg/kg, 100 mg/kg, and 250 mg/kg body weight/day, respectively, for 16 weeks. Then, adipose tissue was immediately collected and used for further analysis.

Isolation of Total RNA

Total RNA was isolated from control and experimental samples using the TRIR kit. Briefly, fresh tissue (100 mg) was homogenized with 1 ml TRIR, followed by centrifugation. The resulting supernatant was

processed with chloroform and isopropanol to isolate RNA pellets, which were then dissolved in autoclaved Milli-Q water.

Quantification of RNA

RNA samples were quantified spectrophotometrically by measuring absorbance at 260/280 nm. The concentration of RNA was calculated based on absorbance values, with a ratio of absorbance at 260/280 nm > 1.8 indicating good quality RNA.

Reverse Transcriptase - Polymerase Chain Reaction (RT-PCR)

RT-PCR was performed using the Eurogentec RT kit. First-strand cDNA synthesis was carried out using oligo dT, dNTPs, and reverse transcriptase, followed by PCR amplification using SYBR green master mix in a real-time PCR system. Primer sequences for target genes (IGF-1 and TNF- α) and β -actin as the invariant control were used [28-29]. Primers: Details of primers used in the present study. IGF-1- FW- 5'-CTGAGCTGGTGGATGCTCT- 3'; RW- 5'-CACTCATCCACAATGCCTGT - 3'; TNF- α - FW - 5' -CAG CGG CCG CAA CAC ATC TCC CTC CGG AAA GGA C - 3' RW - 5' - GAC CGC ACA AGT AGG CAA GAG ATG GCG CCG GCG - 3'; β -actin- FW - 5'-TACAGCTTCACCACCACAGC - 3'; RW- 5'- TCTCCAGGGAGGAAGAGGAT - 3'.

Statistical Analysis

Triplicate analyses of control and treated rat experiments were expressed as mean \pm standard deviation. Statistical analysis was conducted using one-way analysis of variance (ANOVA) followed by Duncan's multiple range test. Results with $p < 0.05$ were considered statistically significant. Graph Pad Prism version 5 was used for analysis.

Results

Impact of Glyphosate on IGF-1 and TNF- α mRNA Expression in Adipose Tissue

The mRNA expression levels of IGF-1 and TNF- α were evaluated via Real-Time PCR to understand the effects of glyphosate exposure. A significant dose-dependent increase ($p < 0.05$) in both IGF-1 and TNF- α expression was observed in glyphosate-treated rats compared to the control group, which displayed no alterations in gene expression. Specifically, as glyphosate dosage increased (50 mg, 100 mg, and 250 mg), TNF- α expression escalated, triggering an inflammatory response. Figure 1 illustrates the impact of glyphosate on IGF-1 mRNA expression, with a clear elevation observed with increasing glyphosate exposure. In brief, IGF-1 mRNA expression was a 0.4-fold

increase in 50 mg glyphosate-treated rats ($p < 0.05$) compared with control while 0.5-fold and 0.7-fold changes were observed in 100 mg, and in 250 mg ($p < 0.05$) glyphosate treated rats respectively (Figure 1). Similarly, the impact of glyphosate on TNF- α mRNA expression (Figure 2) revealed a dose-dependent increase in expression levels, indicating heightened inflammatory response with greater glyphosate exposure. In brief, compared to control, TNF- α mRNA expression was 0.4-fold increase in 50mg glyphosate exposed rats ($p < 0.05$) whereas in 100 and 250 mg glyphosate exposed rats showed a 0.7- and 0.8-fold increase in the mRNA compared to control and 50mg treated rats. This study clearly indicates that glyphosate exposure has diabetogenic potential via the modulation of IGF-1 and TNF- α signaling in the adipose tissue.

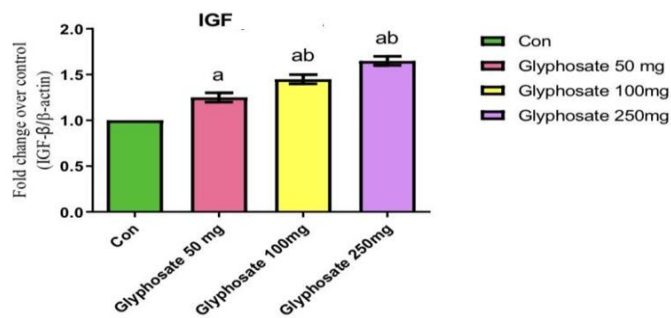


Figure 1. Impact of Glyphosate on mRNA Expression on IGF- 1 In Adipose Tissue of Adult Male Wistar Rats

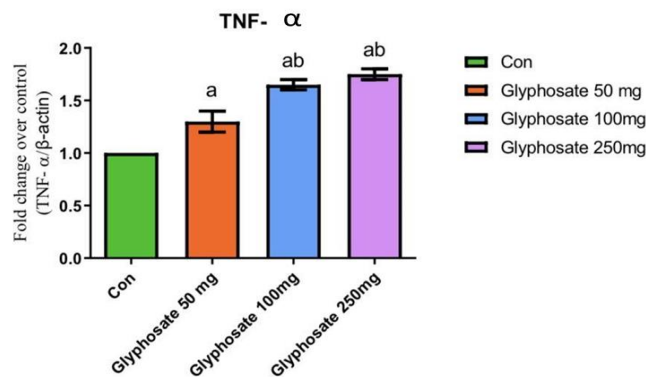


Figure 2. Impact of Glyphosate on mRNA Expression on TNF-alpha. The X-Axis Represents a Controlled Group of Rats in Comparison to Rats Who Are Exposed to Subsequent Doses of Glyphosate

Discussion

This study uncovered significant alterations in IGF-1 and TNF- α gene expression following glyphosate exposure, leading to an inflammatory response associated with potential disorders such as arteriosclerosis in rats [30]. IGF-1, a polypeptide with 70 amino acids, is widely distributed in mammalian tissues. On the other hand, TNF- α , encoded by the human TNF- α gene located on chromosome 6p21.1–21.3, is a cytokine secreted by immune cells implicated in tumour cell necrosis and autoimmune diseases. Dysregulation of TNF- α levels can contribute to various pathological conditions.

TNF- α plays diverse roles in cell survival, signalling, and proliferation, and its aberrant expression is linked to diseases like psoriasis, diabetes, and rheumatoid arthritis [31]. Glyphosate's toxic effects have been implicated in neurodegenerative diseases such as Alzheimer's by increasing TNF- α levels in the brain [7]. Moreover, TNF- α has been associated with ischemic injuries like stroke and can inhibit IGF phosphorylation, contributing to diabetes pathogenesis [32-36].

IGF-1, also known as Insulin-like Growth Factor, exhibited a significant increase upon glyphosate exposure, potentially contributing to diabetes development in rats [32-36]. This finding aligns with previous studies indicating IGF-1's role in bone formation and its therapeutic potential. Additionally, TNF- α expression increased with subsequent glyphosate doses, corroborating its involvement in various diseases and immune responses [37-40].

Further research is warranted to elucidate the mechanisms linking IGF-1 and TNF- α

expression with glyphosate exposure and diabetes progression. [41-45]. Exploring downstream signalling pathways of proinflammatory mechanisms could provide insights into glyphosate's diabetogenic potential and aid in the development of novel therapeutic interventions.

Conclusion

In conclusion, glyphosate exposure exacerbates inflammation, potentially contributing to diabetes progression in adipose tissues by altering IGF-1 and TNF- α expression. Future investigations focusing on proinflammatory transcription factors' protein expression in response to glyphosate exposure are essential to delineate its mechanisms of action in diabetes development.

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Conflict of Interest

The author declares that there is no conflict of interest in the present study.

References

[1] Gardner, J. G., Nelson, G. C., 2008, Herbicides, glyphosate resistance and acute mammalian toxicity: simulating an environmental effect of glyphosate-resistant weeds in the USA. *Pest*

Management Science, 64(4), 470–478. <https://doi.org/10.1002/ps.1497>

[2] de Castilhos Ghisi, N., Zuanazzi, N. R., Fabrin, T. M. C., Oliveira, E. C. 2020, Glyphosate and its toxicology: A scientometric review. *The Science of*

- the Total Environment*, 733, 139359. <https://doi.org/10.1016/j.scitotenv.2020.139359>
- [3] Clair, E., Mesnage, R., Travert, C., Séralini, G. É., 2012, A glyphosate-based herbicide induces necrosis and apoptosis in mature rat testicular cells in vitro, and testosterone decreases at lower levels. *Toxicology in vitro : an international journal published in association with BIBRA*, 26(2), 269–279. <https://doi.org/10.1016/j.tiv.2011.12.009>.
- [4] Gętek-Paszek, M., Całyński, B., Ganczarek-Gamrot, A., Janion, K., Muc-Wierzgoń, M., Nowakowska-Zajdel, E., 2020, Recommendations of the Polish Society of Diabetology and the Lifestyle of Patients with Type 2 Diabetes Mellitus: An Own Research. *Healthcare (Basel, Switzerland)*, 8(4), 504. <https://doi.org/10.3390/healthcare8040504>.
- [5] Wild S. H., 2011, Diabetes, treatments for diabetes and their effect on cancer incidence and mortality: attempts to disentangle the web of associations. *Diabetologia*, 54(7), 1589–1592. <https://doi.org/10.1007/s00125-011-2169-6>
- [6] Thomas, A. G., Holly, J. M., Taylor, F., Miller, V., 1993, Insulin like growth factor-I, insulin like growth factor binding protein-1, and insulin in childhood Crohn's disease. *Gut*, 34(7), 944–947. <https://doi.org/10.1136/gut.34.7.944>
- [7] Parameswaran, N., Patial, S., 2010, Tumor necrosis factor- α signaling in macrophages. *Critical reviews in eukaryotic gene expression*, 20(2), 87–103. <https://doi.org/10.1615/critreveukargeneexpr.v20.i2.10>
- [8] Wu, F., Zhu, J., Li, G., Wang, J., Veeraraghavan, V. P., Krishna Mohan, S., & Zhang, Q. (2019). Biologically Synthesized Green Gold Nanoparticles from Siberian Ginseng Induce Growth-Inhibitory Effect on Melanoma Cells (B16). *Artificial Cells, Nanomedicine, And Biotechnology*, 47(1), 3297–3305. <https://doi.org/10.1080/21691401.2019.1647224>
- [9] Chen, F., Tang, Y., Sun, Y., Veeraraghavan, V. P., Mohan, S. K., Cui, C., 2019, 6-shogaol, an active constituents of ginger prevents UVB radiation mediated inflammation and oxidative stress through modulating Nrf2 signaling in human epidermal keratinocytes (HaCaT cells). *Journal of Photochemistry and Photobiology. B, Biology*, 197, 111518. <https://doi.org/10.1016/j.jphotobiol.2019.111518>
- [10] Li, Z., Veeraraghavan, V. P., Mohan, S. K., Bolla, S. R., Lakshmanan, H., Kumaran, S., Aruni, W., Aladresi, A. A. M., Shair, O. H. M., Alharbi, S. A., Chinnathambi, A., 2020, Apoptotic induction and anti-metastatic activity of eugenol encapsulated chitosan nanopolymer on rat glioma C6 cells via alleviating the MMP signaling pathway. *Journal of Photochemistry and Photobiology. B, Biology*, 203, 111773. <https://doi.org/10.1016/j.jphotobiol.2019.111773>
- [11] Babu, S., Jayaraman, S., 2020, An update on β -sitosterol: A potential herbal nutraceutical for diabetic management. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*, 131, 110702. <https://doi.org/10.1016/j.biopha.2020.110702>
- [12] Malaikolundhan, H., Mookkan, G., Krishnamoorthi, G., Matheswaran, N., Alsawalha, M., Veeraraghavan, V. P., Krishna Mohan, S., Di, A., 2020, Anticarcinogenic effect of gold nanoparticles synthesized from *Albizia lebbek* on HCT-116 colon cancer cell lines. *Artificial cells, Nanomedicine, and Biotechnology*, 48(1), 1206–1213. <https://doi.org/10.1080/21691401.2020.1814313>
- [13] Han, X., Jiang, X., Guo, L., Wang, Y., Veeraraghavan, V. P., Krishna Mohan, S., Wang, Z., Cao, D. 2019, Anticarcinogenic potential of gold nanoparticles synthesized from *Trichosanthes kirilowii* in colon cancer cells through the induction of apoptotic pathway. *Artificial Cells, Nanomedicine, And Biotechnology*, 47(1), 3577–3584. <https://doi.org/10.1080/21691401.2019.1626412>
- [14] Gothai, S., Muniandy, K., Gnanaraj, C., Ibrahim, I. A. A., Shahzad, N., Al-Ghamdi, S. S., Ayoub, N., Veeraraghavan, V. P., Kumar, S. S., Esa, N. M., Arulselvan, P., 2018, Pharmacological insights into antioxidants against colorectal cancer: A detailed review of the possible mechanisms. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*, 107, 1514–

1522. <https://doi.org/10.1016/j.biopha.2018.08.112>
- [15] Veeraraghavan, V.P., Hussain, S., Balakrishna, J.P., Dhawale, L., Kullappan, M., Ambrose, J.M, et al., 2021, A Comprehensive and Critical Review on Ethnopharmacological Importance of Desert Truffles: *Terfezia claveryi*, *Terfezia boudieri*, and *Tirmania nivea*, *Food Reviews International*, 1–20.
- [16] Sathya, S., Ragul, V., Veeraraghavan, V.P., Singh, L., Niyas Ahamed, M.I., 2020, An in vitro study on hexavalent chromium [Cr(VI)] remediation using iron oxide nanoparticles-based beads, *Environmental Nanotechnology, Monitoring & Management*, 2020, 14, 100333.
- [17] Yang, Z., Pu, M., Dong, X., Ji, F., Priya Veeraraghavan, V., Yang, H., 2020, Piperine loaded zinc oxide nanocomposite inhibits the PI3K/AKT/mTOR signaling pathway via attenuating the development of gastric carcinoma: In vitro and in vivo studies. *Arabian Journal of Chemistry*, 13(5), 5501–16.
- [18] Rajendran, P., Alzahrani, A.M., Rengarajan, T., Veeraraghavan, V.P., Krishna Mohan, S., 2020, Consumption of reused vegetable oil intensifies BRCA1 mutations, *Crit Rev Food Sci Nutr*, 27, 1–8.
- [19] Barma, M.D., Muthupandian, I., Samuel, S.R., Amaechi, B.T., 2021, Inhibition of *Streptococcus mutans*, antioxidant property and cytotoxicity of novel nano-zinc oxide varnish. *Arch Oral Biol*, 126, 105132.
- [20] Samuel, S.R., Can 5-year-olds sensibly self-report the impact of developmental enamel defects on their quality of life? *Int J Paediatr Dent*, 31(2), 285–6.
- [21] Samuel SR, Kuduruthullah S, Khair AMB, Shayeb MA, Elkaseh A, Varma SR. Dental pain, parental SARS-CoV-2 fear and distress on quality of life of 2- to 6-year-old children during COVID-19. *Int J Paediatr Dent*. 2021 May;31(3):436–41.
- [22] Tang, Y., Rajendran, P., Veeraraghavan, V.P., Hussain, S., Balakrishna, J.P., Chinnathambi, A., et al. 2020, Osteogenic differentiation and mineralization potential of zinc oxide nanoparticles from *Scutellaria baicalensis* on human osteoblast-like MG-63 cells, *Materials Science and Engineering: C*, 119, 111656.
- [23] Yin Z, Yang Y, Guo T, Veeraraghavan VP, Wang X. Potential chemotherapeutic effect of betalain against human non-small cell lung cancer through PI3K/Akt/mTOR signaling pathway. *Environ Toxicol*. 2021 Jun;36(6):1011–20.
- [24] Veeraraghavan, V. P., Periadurai, N. D., Karunakaran, T., Hussain, S., Surapaneni, K. M., Jiao, X., 2021, Green synthesis of silver nanoparticles from aqueous extract of *Scutellaria barbata* and coating on the cotton fabric for antimicrobial applications and wound healing activity in fibroblast cells (L929), *Saudi Journal of Biological Sciences*, 28(7), 3633–3640.
- [25] Mickymaray, S., Alfaiz, F. A., Paramasivam, A., Veeraraghavan, V. P., Periadurai, N. D., Surapaneni, K. M., Niu, G., 2021, Rhaponticin suppresses osteosarcoma through the inhibition of PI3K-Akt-mTOR pathway. *Saudi Journal of Biological Sciences*, 28(7), 3641–3649.
- [26] Teja, K.V., Ramesh, S., 2020, Is a filled lateral canal – A sign of superiority? *Journal of Dental Sciences*, 15, 562–3.
- [27] Kadanakuppe, S., Hiremath, S., 2016, Social and Behavioural Factors Associated with Dental Caries Experience among Adolescent School Children in Bengaluru City, India, *British Journal of Medicine and Medical Research*. 14, 2016. p. 1–10.
- [28] Garibyan, L., Avashia, N., 2013, Polymerase chain reaction. *The Journal of Investigative Dermatology*, 133(3), 1–4. <https://doi.org/10.1038/jid.2013.1>
- [29] Samadikuchaksaraei, A., 2016, Polymerase Chain Reaction for Biomedical Applications. BoD – Books on Demand, 184.
- [30] Li, J. B., Wang, C. Y., Chen, J. W., Feng, Z. Q., Ma, H. T., 2004, Expression of liver insulin-like growth factor 1 gene and its serum level in patients with diabetes. *World Journal of Gastroenterology*, 10(2), 255–259. <https://doi.org/10.3748/wjg.v10.i2.255>
- [31] Schofield, P.N., 1992, The Insulin-like Growth Factors: Structure and Biological Functions. *Oxford University Press, USA*; 284.
- [32] Bonavida B, Granger G. Tumor Necrosis Factor: Structure, Mechanism of Action, Role in Disease and Therapy. S Karger Ag; 1990. 252 p.

- [33] Jagadheeswari, R., Vishnu Priya, V., Gayathri, R., 2020, Awareness of Vitamin-C Rich Foods Among South Indian Population: A Survey, *Journal of Research in Medical and Dental Science*, 8(7), 330-338.
- [34] Ojastha, B.L., Selvaraj, J., Kavitha, S., Veeraraghavan Vishnu Priya., Gayathri R., 2023, Effect of *Argyrea Nervosa* on The Expression of Growth Factor Signaling In The Skeletal Muscle Of Streptozotocin-Induced Experimental Diabetic Rats. *Journal of Namibian Studies: History Politics Culture*, 33, 5942-5950. <https://doi.org/10.59670/jns.v33i.4474>.
- [35] Selvi, V.T., Devi, R.G., Jothipriya, A. (2020). Prevalence of dental anxiety among the OP patients in Saveetha Dental College. *Drug Invention Today*, 14(1).
- [36] Vishaka, S., Sridevi, G., Selvaraj, J., 2022, An in vitro analysis on the antioxidant and anti-diabetic properties of *Kaempferia galanga* rhizome using different solvent systems. *Journal of Advanced Pharmaceutical Technology & Research*, 13(2), S505–9.
- [37] Ealla KKR, Veeraraghavan VP, Ravula NR, Durga CS, Ramani P, Sahu V, Poola PK, Patil S, Panta P (2022) Silk Hydrogel for Tissue Engineering: A Review. *J Contemp Dent Pract* 23:467–477
- [38] Mithil Vora., Vishnu Priya, V., Selvaraj, J., Gayathri, R., Kavitha, S., 2021, Effect of Lupeol on proinflammatory Markers in Adipose Tissue of High-Fat Diet and Sucrose Induced Type-2 Diabetic Rats. *Journal of Research in Medical and Dental Science*, 9(10), 116-121.
- [39] Sadasivam, P., Ganapathy, D.M., Sasanka, L.K., 2023, Assessment of Depressive Behaviour among the Undergraduate Dental students-A Survey. *Turkish Journal of Physiotherapy and Rehabilitation*, 32, 2.
- [40] Yasothkumar, D., Jayaraman, S., Ramalingam, K., Ramani, P., 2023. In vitro Anti-Inflammatory and Antioxidant Activity of Seed Ethanolic Extract of *Pongamia pinnata*. *Biomedical and Pharmacology Journal*, 16(4).
- [41] Ganesan A, Muthukrishnan A, Veeraraghavan V (2021) Effectiveness of Salivary Glucose in Diagnosing Gestational Diabetes Mellitus. *Contemp Clin Dent* 12:294–300
- [42] Karthik EVG, Priya V (2021) Gayathri. R, Dhanraj Ganapathy. Health Benefits Of *Annona Muricata*-A Review. *Int J Dentistry Oral Sci* 8:2965–2967
- [43] Priya DV, (2020) Knowledge and awareness on HIV/AIDS among college students in A university hospital setting. *Int J Dent Oral Sci* 1182–1186
- [44] Prakash S, Balaji JN, Veeraraghavan VP, Mohan SK (2022) Telehealth: Is It a Post-COVID Reality in Early Diagnosis of Oral Cancer? *J Contemp Dent Pract* 23:1181–1182
- [45] Priya VV, Sankaran K (2023) Aryl hydrocarbon receptor (AhR) as a potential therapeutic target in oral diseases. *World J Dent* 14:1–2